Uni-edit Sample of Level 3 Editing (Biology)

Comment [A1]:

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Material and Methods

Field survey

Manila clams (Nn=30) were collected monthly collected-from a tidal flat off Tokuyama, Yamaguchi Prefecture, western Seto Inland Sea (A-in-Fig. 2, Location A) from June 2004 to May 2005.Collected clams were kept in running seawater for two or three days and then transported to our laboratory in refrigerator overnight. Few clams died during the storage and transportation. Clams were measured at the Sshell length (SL-in; mm), shell-height (SH-in; mm) , shell widthand width (SW-in; mm) was measured prior to removal of .-Ssoft tissue-.was removed from clams and The wet soft tissue weight (WST-in; mg) was then recorded weighted-after removing excess fluid by placing them on filter papers and athe .-cCondition index (CI) was calculated using the following formula: WST/(SL x SH x SW)x1000.

The intensity of *Perkinsus* infection was quantified by conventional Ray's fluid thioglycollate medium (FTM) following-according to Choi et al. (1989) and Almeida et al. (1999). Briefly, the outmost left or right gill leaf was removed from each clam, weighed after removing the excess moisture on filter papers, and subsequently incubated in Ray's FTM medium at 25 °C for one week. Incubated gills were treated –in 2N NaOH at 60 °C until the gills were lysed , mostly within 30 min, and then washed three times in PBS with centrifugation (1600xg, 15 min).

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Resultant pellets were suspended in 1 mL of PBS and a total of 10 μ L of the resuspension was observed-used to calculate the infection load. with an inverted microscope for counting prezoosoprongia. Considering the weight of gill leaves, detection sensitivity of the method was estimated at approximately 10³ cells/g gill tissue. This quantification protocol was employed throughout the present study unless otherwise stated

After removing the outmost gill leaf for examination of infection intensity, the remaining soft tissue was transversally cut in half-into anterior and posterior parts. The eut surface of the posterior part was impregnated on glass slides and the anterior part was fixed in 10% buffered formalin for histological examination. Impregnated tissue preparations were stained using a commercial cytostaining kit (Diff-Quick staining, Symex International Reagents Co. Ltd, Kobe, Japan). For histological examination, a tissue slice of approximately 5 mm thickness was excised out from each of the fixed tissue, embedded in paraffin, sectioned at 5 µm and stained using hematoxylin and eosin. Impregnation preparations and histological sections were observed with a light microscope for examination of sporocysts of trematodes and sexual development of clams, respectively. Sexual development was categorized into 4-four stages: undifferentiated, developing, spawning and spent stages.

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Effects of Perkinsus infection on physiological conditions of clams

The clearance rate, borrowing activity and tolerance against high water temperature were examined together with infection intensity of clams purchased from a local clam farm in Ohno (B-on-Fig. 2, Location B),-). At this location, where the level of *Perkinsus* sp. infection level-was known-high and both *P. olseni* and *P. honshuensis* were present-detected (Takahashi et al., submitted). Clams-were transported in refrigerator overnight to t Clearance rates and tolerance against high water temperature were examined at The Fisheries Laboratory, the-University on-of Tokyo, located on an inlet Hamana Lake, Shizuoka Prefecture (C in Fig. 2, Location C), while borrowing activity was examined at where clearance rates and tolerance against high water temperature were examined, while clam were also transported in refrigerator overnight to our laboratory at the Hongo Campus, the-University of Tokyo, located in Tokyo (D in Fig. 2, Location D), where the borrowing activity was examined.

Clearance rates were measured in 30 clams (SL, 33.2-44.3 mm) with the indirect method in August. Before measurements, clams were acclimatized to-in running seawater at 20°C for 24 hr. For measurements, individual clams were individually placed in a in seawater in a chamber of containing 500 ml of seawater ; and were continuously given commercially cultured diatom, Chaetoceros calcitrans (6.3x10⁴-2.9x10⁵ cells/ml) (Sunculture, Nisshin Marintech, Aichi) continuously through a peristaltic pomp. Diatom densities of the inflow (Cin cells/ml) and outflow (Cout cells/ml) of the chambers-were measured with hemocyte counting chambers. The flow rate (Fr ml/min) of the peristaltic pomp was adjusted between 440-900 ml/h to keep Cout between 10⁴ - 10⁵ cells/ ml. Clearance rates (Cr) were calculated with the following formula: Cr (ml/min) = Fr x (C_{in} - C_{out})/ C_{in} . As the clearance rates was found to became stable-stabilized within 2 hr after-of transferring themclams clams were transferredinto in the chambers in our preliminary experiment, the measurement of Cout and Cin was carried was measured out 5 times every one-hour from 2 hr after transfer of clam into the chambers. After the experiment, infection intensity in the left outmost gill leaf was examined with Ray's FTM.

For examination of the tolerance against high temperature, clams collected from Lake Hamana, which were moderately infected with *Perkinsus*-at a medium level, were compared against also used together with heavily infected clams transported

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from the clam farm located onfrom the Seto Inland Sea in August. Initial infection intensity and condition indexes were examined-recorded for in-30 individual clams from each population using s of clams in each group with Ray's FTM. Seventy six clams each-collected from Lake Hamana and Seto Inland Sea were then from the two groups were-placed in separate baskets placed together withinin a 40 L tank, in through which temperature-controlled flow through seawater was givenpassed. Water temperature was gradually raised-increased from 22 °C to 28 °C in-over a four day period 4 days, and subsequently then maintained at 28 °C for 8 days. During the experiment, Clam survival was recorded twice a day with clams were observed twice a day and dead and moribund or dead clams were being removed from the tank. After the experiment, infection intensity in-of the outmost gill leaf was examined with Ray's FTM.

Borrowing activity was examined in-for 30 clams transported-from the clam farm on-Seto Inland Sea in June. Before the examination, clams were acclimatized at 20°C for one week and fed diatom suspension. For examination, eEach clam was then individually-placed in a 1 L seawater tank with quartzose sand set and placed on the bottom of 100 L recirculating tanks. Water temperature was maintained at 20 °C. Borrowing of clams was observed continuously for <u>-initial-2</u> hrs, then every 30 min for the next 4 hours and once at 24 hours-after the begging of the experiment. The time taken for Timing when each clam shell to completely-disappeared under the sand was recorded. After the experiment, infection intensity in the outmost gill leaf was examined with Ray's FTM.

Statistical analyses

One-way ANOVA followed by Turkey-Kramer HSD test was used for multiple comparisons. Student's t-test was used for comparison between two groups.

Comment [A7]: CHECK: Please specify exactly which comparisons were made using ANOVA. Also indicate if a test for homogeneity of variance was performed before the ANOVA.

Comment [A8]: CHECK: Please specify which two groups were compared using Student's t-test.

Comment [A9]: CHECK: Please specify why the data needed to be transformed. For example, you could write "the intensity data was log-transformed in order to satisfy the assumption of homogeneity of variance".